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Genetic variation in some isoenzyme loci in Scots pine (*Pinus silvestris* L.) populations*

Majority of genetic studies on Scots pine conducted so far was based on an analysis of quantitative and qualitative characters coded by unknown genetic systems. Using biometrical methods a clinal variation was demonstrated for such characters of Scots pine as seed weight, needle colour, time of bud set etc. Langlet (1959) believes that characters having adaptive value are characterized in Scots pine by a clinal variation conditioned by environmental influences. Other authors, e.g. Wright and Bull (1963) believe that the transcontinental range of Scots pine consists of a large number of various genetically distinct races among which a more or less intense gene flow can take place but between which boundaries are definable.

The electrophoretic method of isozyme separation developed since the early seventies permits an expansion of population studies to include several new characters having a precise number of alleles and loci coding them. This method permits also the determination of such population parameters as their uniformity, degree of heterozygosity, and allows the calculation of genetic distances between individual populations basing on frequencies of genes and genotypes of the studied characters.

In the study reported here two isozyme systems have been investigated L-leucinaminopeptidase (3.4.1.1.) and acid phosphatase (3.1.3.2), in the female gametophyte tissue isolated from seeds in a state of rest.

MATERIALS AND METHODS

The studies covered 19 populations of Scots pine. Fourteen were from Poland (Fig. 1), two from Turkey (Culhali and Catak), one from West Germany (Göttingen), one from USSR (Novosibirsk) and one from Hungary (Vas). Seeds of the Turkish populations have been collected

* This work has been supported by grant 10.2. 10.07 from the Polish Academy of Sciences.

from 50 - 60 year old trees. In each of the population a joint seed sample was collected from at least 25 trees. In the remaining 17 populations the seeds were collected from 25 - 35 single trees aged 100 to 140 years. From the Göttingen and Goleniów population seeds were collected from 10 trees only. For calculations of gene frequencies not only the material

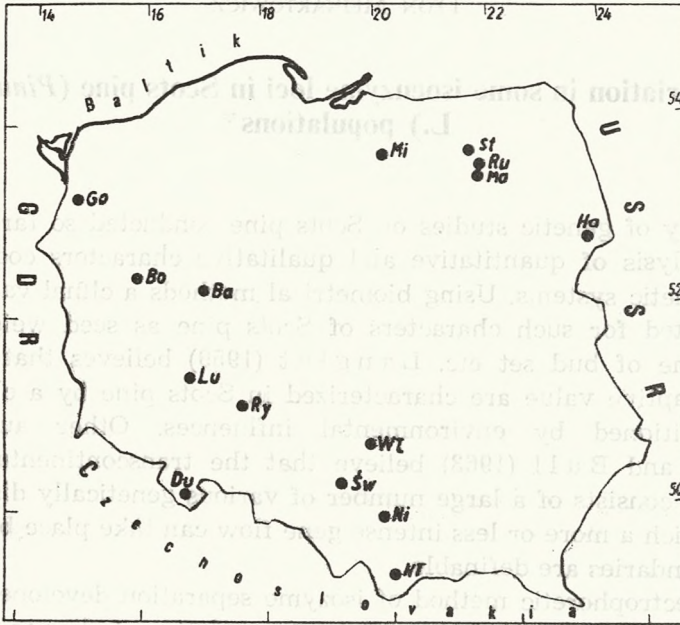


Fig. 1. Map of Poland with location of populations indicated by dots

from single trees was employed but also data from an additional joint sample of 10 trees from each population. The enzymes were extracted from the so called endosperm, that is from the female gametophyte, a haploid tissue, which for this reason is a very convenient material for genetic studies.

The separation of isozymes was performed horizontally on a 12% starch gel in a discontinuous buffer system (Poulik, 1957). For the homogenization of gametophytes in order to isolate LAP (L-leucinamino-peptidase) isozymes a phosphate homogenization buffer pH 7.5 with 5 mM cystein, 5 mM ascorbic acid and 5 mM saccharose was used. For the acid phosphatase (APH) use was made of Tris-Maleic/NaOH, pH 7.4 buffer without any additions. As a substrate for LAP use was made of L-leucyl-beta-naphthylamide hydrochloride and for APH of sodium-alpha-naphthylphosphate (acid). For the visualisation of isozymes on gels after electrophoresis use was made of diazide salts Fast-Black K salt for LAP and Fast Garnet GBC salt for APH. Details of the studied isozymes have been presented in the studies of Bergmann (1973, 1975) on spruce, Mejnartowicz and Bergmann (1975) on larch, and Mej-

nartowicz (1976) on Douglas fir. A genetic and biochemical evaluation of LAP in pine has been given in the works of Białobok et al. (1976), Mejnartowicz (1976), Krzakowa et al. (1977), Rudin (1977), and Salmia and Mikola (1975). A genetic and biochemical analysis of APH has been presented in the work of Mejnartowicz (1978). The interpretation of coding of APH in the female pine gametophyte given by Mejnartowicz et al. (1978) disagrees with the interpretation contained in the work of Krzakowa et al. (1977).

RESULTS

LEUCINE AMINOPEPTIDASE (LAP)

Visualisation of the products of electrophoresis indicated that isoenzymes of LAP occur on gels in two zones, which on the zymograms are designated as LAP-A and LAP-B (Fig. 2). On the basis of studies conducted so far it was established that both zones are coded by two different loci (Mejnartowicz, 1976a; Mejnartowicz et al., 1978; Rudin, 1977).

Locus LAP-A is coded by 5 alleles among which LAP-A1 occurs most commonly in the studied populations, with frequencies from 0.86 in the Lubiń and Duszyniki populations to 1.00 in the Nowy Targ population (Tab. 1). Quite frequently the recessive allele LAP-A4 occurs, attaining a value of 0.14 in the Lubiń population and 0.12 in the Goleniów population. Homozygotic trees for this allele were not found. Least frequent is the allele LAP-A5. It contains a phenotypic pattern in the form of two intensely staining bands. In over 500 studied trees of the European range of Scots pine it was found only twice, and in each of the two Turkish populations it was found at a frequency of 0.03.

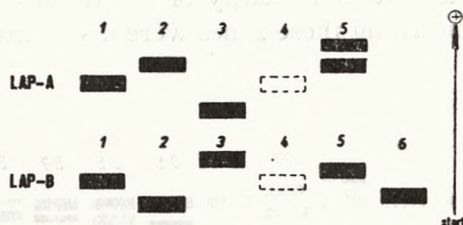


Fig. 2. Schematic drawing of electrophoretic LAP patterns found in extracts of Scots pine macrogametophytes

Locus LAP-B is coded by isozyme molecules of lower electrophoretic mobility. Among 6 observed alleles at this locus the highest frequency of occurrence was found for LAP-B1, attaining 0.97 in the Hajnówka population. The lowest frequency of this allele was found in the Culhali (0.56), Vas (0.63) and Goleniów (0.68) populations in which also a considerable frequency of the allele LAP-B3 was observed (Go — 0.26, Va — 0.17

Table 1

Allele frequencies of the LAP-A and LAP-B loci in seed samples of 19 populations of Scots pine

Population name	LAP-A					LAP-B					
	A1	A2	A3	A4	A5	B1	B2	B3	B4	B5	B6
Goleniów (Go)	0.88	0.0	0.0	0.12	0.0	0.68	0.05	0.26	0.01	0.0	0.0
Babki (Ba)	0.94	0.02	0.0	0.04	0.0	0.83	0.08	0.06	0.0	0.03	0.0
Bolewice (Bo)	0.92	0.0	0.04	0.04	0.0	0.88	0.05	0.05	0.0	0.02	0.0
Göttingen (Gö)	0.89	0.02	0.0	0.09	0.0	0.95	0.0	0.05	0.0	0.0	0.0
Novosibirsk (Ns)	0.99	0.01	0.0	0.0	0.0	0.90	0.02	0.03	0.03	0.02	0.0
Milomłyn (Mi)	0.91	0.0	0.0	0.09	0.0	0.82	0.16	0.02	0.0	0.0	0.0
Strzałowo (St)	0.91	0.0	0.02	0.07	0.0	0.83	0.11	0.06	0.0	0.0	0.0
Duszynki (Du)	0.86	0.0	0.05	0.09	0.0	0.86	0.07	0.07	0.0	0.0	0.0
Świerklaniec (Św)	0.93	0.01	0.0	0.06	0.0	0.85	0.06	0.08	0.0	0.0	0.01
Lubiń (Lu)	0.86	0.0	0.0	0.14	0.0	0.78	0.19	0.03	0.0	0.0	0.0
Rychtal (Ry)	0.98	0.0	0.02	0.0	0.0	0.93	0.03	0.04	0.0	0.0	0.0
Maskulińskie (Ma)	0.79	0.0	0.10	0.11	0.0	0.88	0.01	0.07	0.02	0.02	0.0
Włoszczowa (Wł)	0.99	0.0	0.0	0.01	0.0	0.91	0.08	0.01	0.0	0.0	0.0
Hajnówka (Ha)	0.97	0.0	0.0	0.03	0.0	0.97	0.03	0.0	0.0	0.0	0.0
Nowy Targ (NT)	1.00	0.0	0.0	0.0	0.0	0.91	0.09	0.0	0.0	0.0	0.0
Niepolomice (Ni)	0.93	0.0	0.07	0.0	0.0	0.82	0.02	0.14	0.0	0.02	0.0
Catacik (Ca)	0.97	0.0	0.0	0.0	0.03	0.86	0.12	0.0	0.0	0.02	0.0
Culhali (Cu)	0.97	0.0	0.0	0.0	0.03	0.56	0.0	0.16	0.0	0.0	0.28
Vas (Va)	0.97	0.0	0.0	0.03	0.0	0.63	0.0	0.17	0.07	0.13	0.0

and Cu — 0.16) which in the remaining populations occurs at much lower frequencies was observed for the allele LAP-B6. It occurs abundantly in the Culhali population attaining a value of 0.28 and very rarely in Świerklaniec (0.01). In the remaining populations its occurrence was not recorded.

ACID PHOSPHATASE (APH)

Zymogram of the electrophoretic separation of acid phosphatase is presented in Fig. 3. It consists of four zones designated as APH-A, APH-B, APH-C and APH-D. In view of the low activity of enzymes and insufficient reproducibility of results in zone A, C and D the results obtained concerning these zones were disregarded.

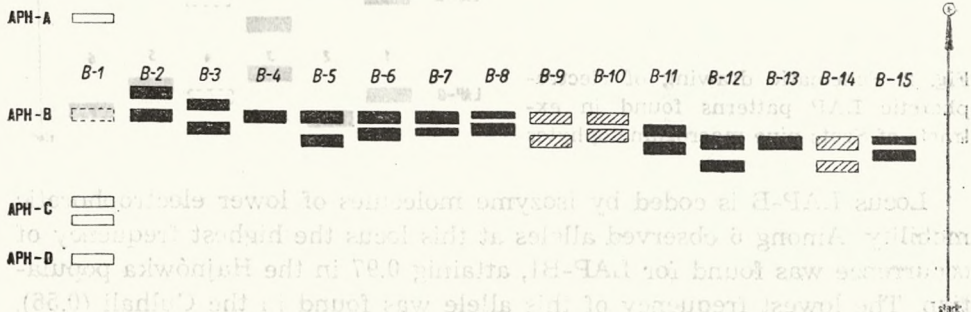


Fig. 3. Schematic drawing showing the electrophoretic variants in the one APH-B locus found in extracts of Scots pine gametophytes

Table 2

Allele frequencies of the APH-B locus in seed samples of 19 populations of Scots pine

Population name	APH-B														
	B1	B2	B3	B4	B5	B6	B7	B8	B9	B10	B11	B12	B13	B14	B15
Goleniów (Go)	0.0	0.0	0.05	0.05	0.70	0.0	0.05	0.0	0.0	0.05	0.0	0.10	0.0	0.0	0.0
Babki (Ba)	0.08	0.0	0.06	0.08	0.62	0.02	0.02	0.0	0.04	0.0	0.0	0.04	0.02	0.02	0.0
Bolevice (Bo)	0.03	0.0	0.0	0.03	0.58	0.13	0.0	0.03	0.07	0.07	0.0	0.03	0.0	0.03	0.0
Göttingen (Gö)	0.0	0.0	0.0	0.0	0.70	0.20	0.0	0.0	0.10	0.0	0.0	0.0	0.0	0.0	0.0
Novosibirsk (Ns)	0.0	0.0	0.0	0.10	0.50	0.06	0.0	0.0	0.0	0.06	0.0	0.26	0.02	0.0	0.0
Miłomłyn (Mi)	0.0	0.0	0.0	0.0	0.58	0.10	0.0	0.03	0.13	0.05	0.03	0.05	0.0	0.0	0.03
Strzałowo (St)	0.0	0.0	0.0	0.02	0.57	0.04	0.02	0.02	0.11	0.10	0.04	0.02	0.02	0.0	0.0
Duszniki (Du)	0.0	0.0	0.02	0.0	0.68	0.0	0.0	0.16	0.02	0.05	0.0	0.0	0.05	0.0	0.02
Świerkianiec (Sw)	0.0	0.0	0.02	0.02	0.66	0.07	0.08	0.02	0.0	0.15	0.0	0.0	0.0	0.0	0.0
Lubiń (Lu)	0.0	0.0	0.0	0.0	0.68	0.0	0.07	0.0	0.0	0.25	0.0	0.0	0.0	0.0	0.0
Rychtal (Ry)	0.0	0.0	0.0	0.0	0.77	0.0	0.0	0.0	0.04	0.19	0.0	0.0	0.0	0.0	0.0
Maskulińskie (Ma)	0.0	0.0	0.0	0.0	0.75	0.0	0.0	0.0	0.03	0.19	0.0	0.03	0.0	0.0	0.0
Włoszczowa (Wl)	0.0	0.0	0.0	0.0	0.75	0.04	0.13	0.0	0.04	0.04	0.0	0.0	0.0	0.0	0.0
Hajnówka (Ha)	0.05	0.0	0.05	0.05	0.60	0.0	0.20	0.0	0.0	0.05	0.0	0.0	0.0	0.0	0.0
Nowy Targ (NT)	0.0	0.0	0.0	0.05	0.60	0.0	0.30	0.0	0.0	0.05	0.0	0.0	0.0	0.0	0.0
Niepołomice (Ni)	0.0	0.0	0.0	0.20	0.40	0.20	0.20	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
Catacik (Ca)	0.01	0.0	0.0	0.57	0.0	0.25	0.0	0.02	0.01	0.0	0.02	0.02	0.10	0.0	0.0
Culhali (Cu)	0.0	0.02	0.06	0.05	0.0	0.84	0.0	0.0	0.0	0.0	0.0	0.01	0.02	0.0	0.0
Vas (Va)	0.13	0.0	0.13	0.57	0.07	0.07	0.0	0.0	0.0	0.03	0.0	0.0	0.0	0.0	0.0

Zone APH-B accumulates the highest quantities of enzyme, of the highest activity and at the same time the results are best reproducible. As was shown in earlier studies this zone is coded by one locus henceforth referred to as locus APH-B (Mejnartowicz, 1978). This locus is coded by at least 15 electrophoretic alleles having a phenotypic form of one or two bands. Allele APH-B1 is the so called „zero allele” or recessive on which codes an enzymatically inactive protein. It occurs in the studied pine populations rather rarely compared with the frequency of occurrence of recessive alleles coding LAP loci. The highest frequency of 0.13 was observed for APH-B1 in the Vas population than 0.08 in Babki 0.03 in Bolevice and 0.01 in Catacik (Tab. 2). However the least frequent allele is the APH-B2 one the appearance of which was observed only in the Culhali population where it had a frequency of 0.02 (Tab. 2).

Many of the electrophoretic alleles presented in the zymogram (Fig. 3) have low frequencies of occurrence and this in a low number of populations. Besides the mentioned above APH-B1 and APH-B2 this group includes alleles APH-B11, APH-B13, APH-B14 and APH-B15. The frequency of these alleles does not exceed a value of 0.10. Some others alleles of APH such as APH-B12 appear with large frequencies only in one population (Novosibirsk 0.26) while in other they do not appear at all or with a very low frequency. Similarly APH-B8 in Duszniki attains a frequency of 0.16 while in the remaining five populations in which its presence was recorded the frequency did not exceed 0.03 and in the remaining 13 populations it was nil (Tab. 2).

The greatest number of acid phosphatase enzymes in almost all popula-

tions is coded by allele APH-B5 except for the Turkish populations in which this allele was not observed and the Hungarian population Vas in which its frequency was only 0.07. In these populations on the other hand high frequencies of other alleles were observed, namely of APH-B4 (0.57 in populations Catacik and Vas) and APH-B6 having a frequency of 0.84 in the Culhali population (Tab. 2).

HETEROZYGOSITY OF SCOTS PINE POPULATIONS

Making use of the formulae suggested by Nei (1972) it is possible to establish the degree of heterozygosity h at each of studied loci and to calculate the mean heterozygosity H for all the populations jointly:

$$\hat{H} = \sum_{k=1}^r h_k / r,$$

where $h_k = 1 - j_x$, $j_x = \sum x_i^2$,

r — number of loci,

h_k — degree of heterozygosity at locus k ,

x_i — frequency of i -th allele in the population.

Among the 19 investigated populations mean heterozygosity lied within the limits from $\hat{H}_{\min} = 0.1804$ to $\hat{H}_{\max} = 0.4137$. Highest heterozygosity was observed in the Hungarian population Vas ($\hat{H} = 0.4137$) and in the Polish population from Goleniów where $\hat{H} = 0.3895$ and Niepołomicé with a mean heterozygosity $\hat{H} = 0.3858$. The high heterozygosity of the Vas population

Table 3

Heterozygosity at three isoenzyme-loci (LAP-A; LAP-B; APH-B) in nineteen populations of *Pinus silvestris*

Population	Loci			\hat{H}
	LAP-A	LAP-B	APH-B	
Goleniów (Go)	0.2112	0.4674	0.4900	0.3895
Babki (Ba)	0.1144	0.3002	0.5944	0.3663
Belewiec (Bo)	0.1504	0.2202	0.6324	0.3343
Göttingen (Gö)	0.1994	0.0950	0.4600	0.2515
Novosybirsk (Ns)	0.0198	0.1874	0.6648	0.2907
Miłomłyn (Mi)	0.1638	0.3016	0.6290	0.3648
Strzałowo (St)	0.1666	0.2954	0.6462	0.3694
Duszynki (Du)	0.2498	0.2506	0.5058	0.3354
Świerkianiec (Św)	0.1314	0.2674	0.5294	0.3094
Lubiń (Lu)	0.2408	0.3546	0.4702	0.3552
Rychtal (Ry)	0.0392	0.1326	0.3694	0.1804
Maskulińskie (Ma)	0.3538	0.2198	0.3996	0.3244
Włoszczowa (Wl)	0.0198	0.1654	0.4158	0.2003
Hajnowka (Ha)	0.0582	0.0582	0.5900	0.2355
Nowy Targ (NT)	0.000	0.1638	0.5450	0.2363
Niepołomicé (Ni)	0.1302	0.3072	0.7200	0.3858
Catacik (Ca)	0.0582	0.2456	0.6012	0.3017
Culhali (Cu)	0.0582	0.5824	0.2874	0.3093
Vas (Va)	0.0582	0.5524	0.6306	0.4137

is the result of a very high heterozygosity of one locus LAP-B while in Niepołomice high heterozygosity results from high heterozygosity in locus APH-B in which four alleles APH-B4, B5, B6, and B7 occur with high frequencies. In the Goleniów populations all three studied loci have a high value of h (Tab. 3).

The population from Rychtal in central Poland has a low heterozygosity of $\bar{H}=0.1804$. In this population in the majority of studied trees all three loci are to a large degree monomorphic. Similar is the case with genetic variability within the population from Włoszczowa, where $\bar{H}=0.2003$.

Some of the studied populations have only one monomorphic locus as for example Novosibirsk, where h in locus LAP-A is 0.0198 while at another locus APH-B it is high $h=0.6648$ indicating a high polymorphism of that locus.

INTER-POPULATION VARIABILITY

While the genetic variability within populations may be evaluated by the mean heterozygosity per locus, differences between two populations are evaluated on the basis of genetic distances D , which were calculated here according to Nei (1972) by the formula:

$$D = -\log_e I$$

and where I is the normalized identity of genes computed by

$$I = \frac{J_{XY}}{\sqrt{J_X J_Y}}$$

in which J_X and J_Y are the average homozygosity per locus in populations X and Y respectively and J_{XY} is the average identity of genes between X and Y. J_X is calculated as:

$$J_X = \sum_{k=1}^r \sum x_i^2,$$

$$J_Y = \sum_{k=1}^r \sum y_i^2,$$

$$J_{XY} = \frac{\sum_{k=1}^r \sum x_i y_i}{r}.$$

It is a measure of similarity between populations and it served to draw a phenogram presented in Fig. 4 on which all the Polish populations except Niepołomice and Goleniów constitute a homogenous group of populations which are closely related to the Lower Saxony population from

Table 4

Coefficients of genic identity (I) above the diagonal and Nei's genetic distance (D -value, Nei, 1972) below the diagonal between 19 populations of Scots pine. Abbreviations identifying samples are as indicated on the map, Fig. 1.

Population	Go	Bo	Mi	Ma	St	Lu	Du	Św	Wi
Goleniów (Go)	—	.968	.962	.974	.971	.960	.971	.977	.969
Bolevice (Bo)	.032	—	.992	.982	.992	.969	.981	.989	.986
Milomłyn (Mi)	.038	.008	—	.975	.994	.974	.979	.980	.983
Maskulińskie (Ma)	.026	.018	.025	—	.987	.984	.983	.991	.981
Strzałowo (St)	.029	.008	.006	.013	—	.979	.985	.990	.988
Lubiń (Lu)	.040	.031	.026	.016	.021	—	.971	.988	.976
Duszniaki (Du)	.029	.019	.021	.017	.015	.029	—	.984	.982
Świerklaniec (Św)	.023	.012	.020	.009	.010	.012	.016	—	.992
Włoszczowa (Wi)	.031	.014	.017	.019	.012	.024	.018	.008	—
Niepołomice (Ni)	.082	.064	.088	.099	.083	.107	.099	.065	.066
Hajnówka (Ha)	.038	.030	.039	.030	.028	.034	.032	.016	.008
Nowy Targ (NT)	.058	.048	.053	.049	.042	.042	.049	.027	.016
Rychtal (Ry)	.031	.014	.021	.006	.010	.015	.017	.006	.012
Vas (Va)	.328	.331	.375	.368	.355	.383	.371	.345	.426
Culhali (Cu)	.459	.389	.397	.475	.435	.489	.468	.409	.443
Catacik (Ca)	.411	.344	.372	.419	.372	.415	.402	.377	.396
Novosybirsk (Ns)	.046	.037	.046	.047	.046	.067	.057	.047	.049
Babki (Ba)	.022	.013	.019	.024	.015	.035	.019	.017	.013
Göttingen (Gö)	.039	.007	.013	.028	.018	.048	.028	.028	.018

Table 4 continued

Population	Ni	Ha	NT	Ry	Va	Cu	Ca	Ns	Ba	Gö
Goleniów (Go)	.921	.962	.946	.969	.720	.632	.663	.955	.978	.961
Bolevice (Bo)	.938	.970	.953	.986	.718	.678	.709	.963	.987	.993
Milomłyn (Mi)	.916	.961	.948	.979	.687	.672	.689	.955	.981	.987
Maskulińskie (Ma)	.906	.970	.952	.994	.692	.622	.658	.954	.976	.972
Strzałowo (St)	.920	.972	.959	.990	.701	.647	.689	.955	.985	.982
Lubiń (Lu)	.898	.966	.959	.985	.682	.613	.660	.935	.965	.953
Duszniaki (Du)	.906	.968	.952	.983	.690	.626	.669	.945	.981	.972
Świerklaniec (Św)	.937	.984	.973	.994	.708	.664	.686	.954	.983	.972
Włoszczowa (Wi)	.936	.992	.984	.988	.653	.639	.673	.952	.987	.982
Niepołomice (Ni)	—	.946	.950	.904	.819	.763	.822	.915	.932	.936
Hajnówka (Ha)	.055	—	.994	.975	.720	.625	.688	.943	.981	.963
Nowy Targ (NT)	.051	.006	—	.959	.704	.625	.689	.928	.962	.945
Rychtal (Ry)	.101	.025	.042	—	.691	.625	.664	.954	.980	.975
Vas (Va)	.200	.328	.351	.370	—	.688	.945	.769	.747	.697
Culhali (Cu)	.270	.470	.470	.470	.374	—	.770	.567	.612	.714
Catacik (Ca)	.196	.374	.372	.409	.057	.261	—	.736	.710	.698
Novosybirsk (Ns)	.090	.059	.075	.047	.263	.567	.306	—	.941	.948
Babki (Ba)	.070	.019	.038	.020	.292	.491	.342	.061	—	.979
Göttingen (Gö)	.066	.037	.057	.025	.361	.337	.360	.053	.021	—

Göttingen (which is very similar to Bolevice). The Turkish populations are clearly distinct. They have a high genetic distance between each other and with all the other populations (Fig. 4). They are related to the Hungarian Vas population. Of intermediate nature between the Turkish-Hungarian group and the Polish group is the Niepołomice population from southern Poland. The population from Goleniów is a pine seed stand with a high degree of intrapopulation variability, having also on the phenogram a somewhat independent position (Fig. 4).

DISCUSSION

The results of studies presented here on the frequency of occurrence of alleles in pine populations indicates that there are considerable differences in the heterozygosity of populations depending on the studied locus. Much more important is the information about the heterozygosity of populations than about the frequency of occurrence of any particular allele. Willis (1973) suggests that the greater adaptability of populations

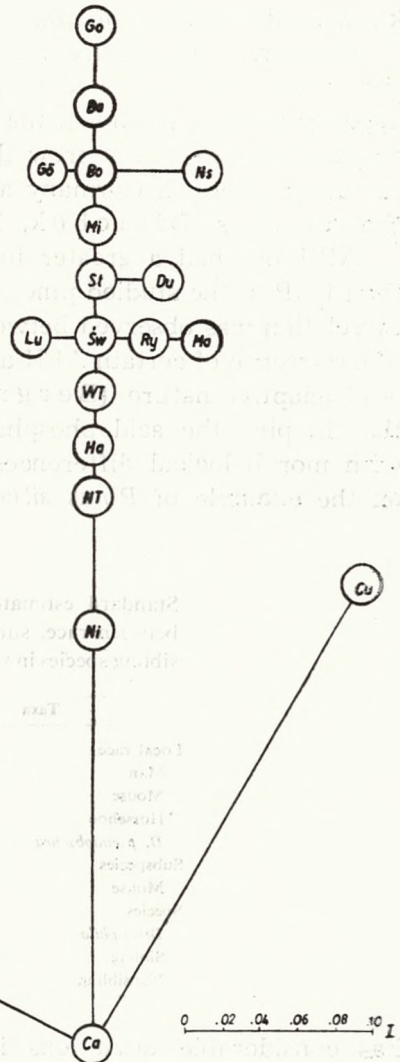


Fig. 4. Phenogram of genic identity based on Nei's I-value for 19 populations samples of Scots pine, calculated jointly for frequencies of APH and LAP

is associated with a greater heterozygosity of enzymes per individual regardless of the individual enzymatic alleles present in the functioning polymorphism. An evaluation of the known part of heterozygosity in populations of Scots pine in Europe encounters difficulties resulting from

the fact that in the majority of existing stands the populations are of artificial origin.

In introduced spruce populations in Sweden a high degree of heterozygosity was observed (Lundkvist and Rudin, 1977). Similarly the undoubtedly introduced Vas population in Hungary has had a high degree of heterozygosity. On the other hand the certainly indigenous populations from Miłomłyn (the famous Tabórz pine race), Maskulińskie and Strzałowo are also highly heterozygotic. It is possible that the well known adaptability of pine from this region (Przybylski, 1972; Troeger, 1960) results from this high heterozygosity. It is also possible that human activity is to some extent responsible for this high heterozygosity since it is known that even in this region where phenotypically magnificent pine races grow there have been introductions of seed from far away places in Germany and France which differ substantially from the local races (Białobok, 1973).

APH has had a greater influence on the measure of heterozygosity than LAP in the studied pine populations. In spruce populations a distinct correlation was observed between ecological conditions and the frequency of occurrence of certain APH alleles. This would indicate that this enzyme is of adaptive nature (Bergmann, 1978). However it does not appear that in pine the acid phosphatase isozymes are in any way associated with morphological differences in pine populations. This can be judged on the example of *Pinus silvestris* f. *conglomerata*. In this form which

Table 5

Standard estimates (*D*) of net codon differences between race, subspecies, sibling species, and non sibling species in various organisms (after Nei, 1973)

Taxa	<i>D</i>
Local races	
Man	0.005 - 0.02
Mouse	0.002 - 0.03
Horsehoe	0.002 - 0.04
<i>D. pseudobscura</i>	0.003 - 0.13
Subspecies	
Mouse	0.13 - 0.20
Species	
<i>Drosophila</i>	
Sibling	0.18 - 1.54
Nonsibling	1.3 - 2.54

has considerable deviations in morphology from the type in having a broom like habit, cones in abundant clusters etc. the pattern of APH and LAP isozymes was quite normal (Mejnartowicz, 1979).

Though on the basis of studies conducted so far it is not possible to draw far reaching conclusions, it is perhaps worth noting that the value

of the genetic distance between populations in the north-south direction is much greater than in the east-west direction. There is an exceptionally great distance between the Turkish and Hungarian populations on the one hand and the remaining European populations on the other, the southern Polish population from Niepołomice being intermediate. There are suggestions that the Niepołomice stand are of Austrian origin. The value of the genetic distances between European populations and the southern ones are 0.196 - 0.415 for Catacik, 0.270 - 0.489 for Culhali and 0.222 - 0.426 for Vas. These values indicate distances greater than those usually observed between races within one species (Tab. 5).

CONCLUSIONS

1. In the studied populations of Scots pine occurrence of five alleles coding locus LAP-A and six alleles coding locus LAP-B were observed. The greatest frequencies were observed for LAP-A1 and LAP-B1 and the majority of studied genotypes were of the type LAP-A1A1B1B1.

2. Much greater genetic polymorphism was observed in the studied locus APH-B coding acid phosphatase. This locus is coded by 15 alleles. In European populations the allele APH-B5 was most common.

3. In all populations except Culhali from Turkey the heterozygosity of trees in locus APH-B is much greater than in the LAP-A and LAP-B loci.

4. A large number of heterozygotes was found with the recessive allele LAP-A4 while heterozygotes having the recessive alleles LAP-B4 and APH-B1 are very rare in populations of Scots pine.

5. Considerable differences were found in the genetic structure of populations between the southern part of range, represented by pine from Turkey and from Hungary (and to some extent the southern Polish population from Niepołomice) and the other populations studied.

6. The rather low genetic distance between the Polish populations and the populations from Göttingen and Novosibirsk indicate that there is a much lower variability in Scots pine along the East-West axis than along the North-South one.

SUMMARY

Female gametophytes from seeds of 19 Scots pine (*Pinus silvestris*) from Poland, FRG, USSR, Turkey and Hungary were analysed for the content of leucine aminopeptidase (LAP) and acid phosphatase (APH)

isozymes. A considerable variability of APH enzyme was observed, with 15 alleles coding the APH-B locus. There were two LAP loci, LAP-A coded by 5 alleles and LAP-B coded by 6 alleles. A considerable genetic distance was observed between the Turkish and Hungarian populations. These populations from the southern part of the range differ substantially from the other European ones, the Niepołomice population from southern Poland being intermediate. The variability of Scots pine proved to be much greater along the N-S axis than along the W-E axis. The populations from the Mazury region known for their great adaptability are characterized by considerable heterozygosity.

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LEON MEJNARTOWICZ

Zmienność genetyczna izoenzymów w niektórych loci w populacjach sosny zwyczajnej (*Pinus silvestris* L.)

Streszczenie

Badania izoenzymów leucyloaminopeptydazy (LAP) i kwaśnej fosfatazy wykonano na endospermach nasion zebranych w 19 populacjach sosny zwyczajnej z Polski, RFN, ZSRR oraz Turcji i Węgier. Stwierdzono dużą zmienność enzymu APH, który w badanym locus APH-B kodowany był w postaci 15 alleli. LAP kodowana była w 2 loci, z których LAP-A zawierał 5 alleli, a ALP-B sześć alleli. Stwierdzono znaczny dystans genetyczny między populacjami tureckimi i populacją węgierską. Populacje te, z południowej części zasięgu znacznie różnią się od pozostałych populacji europejskich, łącząc się z nimi przez południowopolską populację Niepołomice. Znacznie większa okazała się zmienność sosny zwyczajnej w kierunku południkowym niż równoleżnikowym.

Znane ze swej zdolności adaptacyjnej populacje sosen mazurskich charakteryzują się dużą zmiennością wewnątrzpopulacyjną, wyrażającą się wysokim stopniem heterozygotyczności tych populacji.

ЛЕОН МЕЙНАРТОВИЧ

*Генетическая изменчивость некоторых изоферментных локусов
в популяциях сосны обыкновенной (Pinus silvestris L.)*

Резюме

В работе представлены исследования генетической изменчивости изоферментов лейцинаминопептидазы (LAP) и кислой фосфатазы (АРН) в эндоспермах семян сосны обыкновенной из 19 популяций с территории Польши, ФРГ, СССР, Турции и Венгрии. В результате исследований была установлена большая изменчивость фермента АРН, который в исследуемом локусе АРН-В кодировался 15 аллелями. LAP кодировался 2 локусами, из которых LAP-A содержал 5 аллелей, а LAP-B — 6 аллелей. Констатирована большая генетическая разница между турецкими и венгерской популяциями. Эти популяции, с южной части ареала распространения сосны обыкновенной, отличаются от остальных европейских популяций, соединяясь с ними через южнопольскую популяцию Неполомице. Изменчивость сосны обыкновенной в меридианном направлении оказалась значительно большей, чем в параллельном.

Известные своей адаптационной способностью популяции мазурских сосен характеризовались большой внутрипопуляционной изменчивостью, что выражалось большой степенью гетерозиготности этих популяций.